SO-C101 induces NK cell cytotoxicity and potentiates antibody-dependent cell cytotoxicity and anti-tumor activity

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Introduction

SO-C101 is a superagonist fusion protein of interleukin (IL)-15 and the IL-15 receptor α (IL-15R α) sushi⁺ domain, representing a promising clinical candidate for the treatment of cancer. SO-C101 specifically stimulates natural killer (NK) cells and memory CD8⁺ T cells with no significant expansion and activation of regulatory T cell compartment. Human NK cell proliferation, the expression of NK cell receptors and ADCC activity of human PBMC after stimulation with SO-C101 in vitro in combination with monoclonal antibodies were detected by flow cytometry. The anti-tumor efficacy of SO-C101 in combination with Daratumumab was assessed in a multiple myeloma xenograft mouse model in vivo. In this study, we show that SO-C101 induced proliferation and expansion of both major subsets of human NK cells, CD56^{bright}CD16⁻ and CD56^{dim}CD16⁺. Furthermore, SO-C101 induced expression of the cytotoxic receptors NKp30 and NKG2D whereas no upregulation of the inhibitory receptors CD158a, CD158b and NKG2A was detected. Both NK cell subsets activated by SO-C101 exhibited cytotoxicity towards cancer cells in vitro. Using human PBMCs, we show that SO-C101 potentiated killing of tumor cells induced by several clinically approved therapeutic monoclonal antibodies such as Cetuximab, Daratumumab and Obinutuzumab in vitro. SO-C101 and Daratumumab monotherapy treatment inhibited tumor growth in vivo, however, their combination showed the strongest anti-tumor efficacy. Specifically, sequential administration of Daratumumab, followed by SO-C101 promoted complete tumor regression, compared to partial anti-tumor responses induced by the respective monotherapies. SO-C101 augments the anti-tumor activity of therapeutic antibodies by increasing NK cells mediated antibody-dependent cell cytotoxicity. These results support the evaluation of SO-C101 in combination with monoclonal therapeutic antibodies in clinical studies.



• SO-C101 (211 amino acids) is a fusion protein that consists of the N-terminal sushi* domain of the human IL-15R α (77 amino acids) covalently coupled via a non-immunogenic linker of Glycine-Serine residues (20 amino acids) to the mature IL-15 sequence (114 amino acids) (Mortier et al., 2006).

SO-C101 acts as a selective and potent agonist of the IL-15 pathway through IL-15R β y thereby inducing proliferation and activation of memory CD8⁺ T cells, NK cells, $\gamma\delta$ T cells and NKT cells *in vitro* and in vivo and exerts significantly increased anti-tumor efficacy in various mouse cancer models over IL-15 (Bessard et al., 2009; Desbois et al., 2016)

Figure 1



Figure 1. Human PBMCs were incubated with the indicated concentrations of SO-C101 for 7 days (NT - non-treated) and NK cells (CD3⁻CD56⁺) were analyzed by flow cytometry. (A, B) SO-C101 stimulates and expands human NK cells and their subpopulations CD56^{dim}CD16⁺ and CD56^{bright}CD16⁻ in a concentration-dependent manner *in vitro*. NK cell proliferation was detected as the percentage of Ki67⁺ cells (C) SO-C101 induces the expression of activation receptors (DNAM-1, NKp30 and NKG2D) on NK cells without inducing the expression of the inhibitory receptors (CD158a, CD158b and NKG2A) after 7 days of stimulation in vitro. Data represent mean + SEM from 4 healthy donors.

Figure 2

SO-C101 stimulates cytotoxic activity of human NK cells in vitro



Figure 2. The cytotoxic activity of human NK cells induced by SO-C101 was analyzed by flow cytometry using LAMP-1 staining as a marker of cytotoxic degranulation, intracellular IFN-γ staining and detection of dead tumor cells (DAPI⁺). (A) NK cells and their both subsets CD56^{dim}CD16⁺ and CD56^{bright}CD16⁻ exhibit cytotoxic activity towards K562 tumor cells. (B) Representative dotplots of LAMP-1 and IFN-γ staining in NK cells are shown (NT - non-treated; 1 nM SO-C101 + K562 - human PBMC treated with SO-C101 and co-cultivated with K562 target cells; 1 nM SO-C101 human PBMC treated with SO-C101 without K562 co-cultivation). (C) The percentage of dead DAPI+ K562 tumor cells was determined by flow cytometry after 6 h treatment. Data represent mean ± SEM from 4 healthy donors.

Figure 3

SO-C101 enhances the activity of therapeutic antibodies in human PBMC killing assay in vitro



Figure 3. SO-C101 potentiates the activity of Daratumumab, Obinutuzumab and Cetuximab to kill tumor cells in a concomitant (A) and sequential (B) treatment setting in human PBMC killing assay in vitro. Percentage of cytotoxicity was determined by a lactate dehydrogenase release assay (Cytotoxicity Detection Kit). (A) Concomitant treatment. Human PBMCs were added to the target tumor cells together with 1 nM SO-C101 and the therapeutic antibodies at the indicated concentrations and incubated for 20 h. E:T ratio was 25:1 for Daudi (Daratumumab, Obinutuzumab) and 150:1 for SCC25 (Cetuximab) cells, respectively. (B) Sequential treatment. Human PBMCs were first stimulated with 0.1 nM SO-C101 for 2 days (SO-C101-PBMC) and then added to the target tumor cells together with therapeutic antibodies at the indicated concentrations and incubated for 24 h. E:T ratio was 15:1 for Daudi and 100:1 for SCC25, respectively. Data represent mean ± SEM from 3 healthy donors. **P < 0.0021, ***P < 0.0002, Wilcoxon signed rank test. (NT - non-treated).





No K562



Figure 4

Figure 4. SO-C101 stimulation of human NK cells (A) and PBMCs (B, C) potentiates antibody-dependent cell cytotoxicity (ADCC) of Daratumumab. (A) Isolated human NK cells were treated with SO-C101 (0.1 nM, 3 days) and then incubated with Daratumumab and Daudi tumor cells (E: T 10:1) for 4 h. Cytotoxicity was determined by a lactate dehydrogenase release assay (B) Human PBMCs were treated with SO-C101 (0.1 nM, 2 days) and then incubated with Daratumumab and Daudi cells (E:T 1:1) for 4 h. Tumor cell death (DAPI⁺) was detected by flow cytometry. (C) Furthermore the cytotoxic activity of human NK cells was determined by LAMP-1 staining as a marker of cytotoxic degranulation. Representative dot plots are shown, where Daratumumab was used at 0.1 nM. Data represent mean ± SEM from 6 (A) or 3 (B, C) healthy donors. ****P < 0.0001, ***P < 0.0002, Wilcoxon signed rank test.

Figure 5

SO-C101 potentiates therapeutic activity of Daratumumab in vivo



Figure 5. (A) SO-C101 and Daratumumab treatment decreases the tumor growth in human multiple myeloma Xenograft model (RPMI8226) in CB17 SCID mice. Mice were treated s.c with SO-C101 at 1 mg/kg and i.p. with Daratumumab at 20 mg/kg according to the schemes. (B) Co-administration of SO-C101 (1 mg/kg, day 7-10) and Daratumumab (20 mg/kg, day 0) demonstrated the most potent anti-tumor activity with tumor growth inhibition (TGI) value of 100 %. Data are presented as mean + SEM from 8 mice.

Conclusions

- SO-C101 expands and activates both subsets of human NK cells in vitro which is accompanied by the expression of the activation receptors
- SO-C101 stimulates the cytotoxic activity of human NK cells towards tumor cells in vitro
- · SO-C101 enhances the activity of Daratumumab, Obinutuzumab and Cetuximab to kill tumor cells in human PBMC killing assay in vitro.
- SO-C101 potentiates antibody dependent cell cytotoxicity (ADCC) of Daratumumab in vitro.
- SO-C101 synergizes with Daratumumab in anti-tumor efficacy in vivo.

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